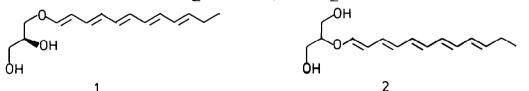
SYNTHESIS OF RACEMIC FECAPENTAENE-12. A POTENT MUTAGEN FROM HUMAN FECES, AND ITS REGIOISOMER

A. A. Leslie Gunatilaka. Nobuhiro Hirai. David G. I. Kingston*

Department of Chemistry Virginia Polytechnic Institute and State University Blacksburg, VA 24061

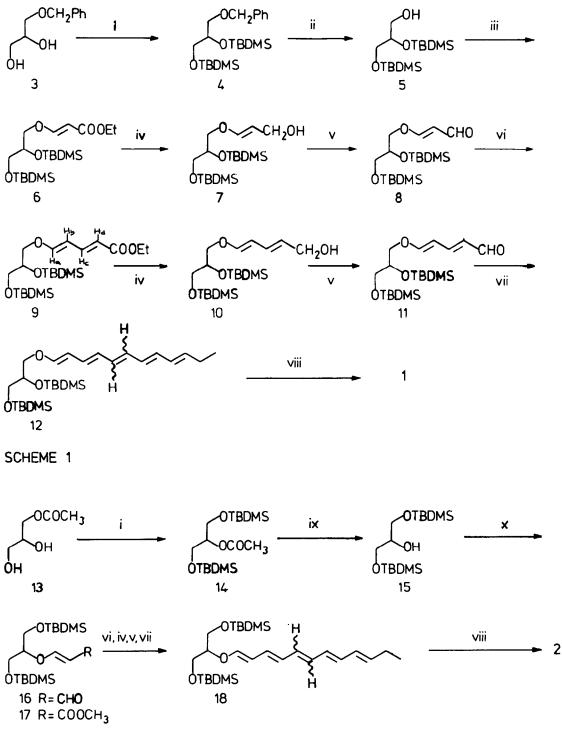
Summary: Racemic fecapentaene-12 [3-(1,3,5,7,9-dodecapentaenyloxy)-1,2-propanediol (1)] and its regioisomer 2-(1,3,5,7,9-dodecapentaenyloxy)-1,3-propanediol (2) have been synthesized. The latter compound is comparably mutagenic to 1.

Recently we elucidated the structure of a potent mutagen produced by colonic bacteria in persons at high risk for colon cancer as the unusual glycerol enol ether 1.¹ Similar conclusions were reached independently by Bruce, who suggested the name fecapentaene for this class of comnounds.² and we have adopted his nomenclature. In this communication we report the first synthesis of racemic fecapentaene-12 (1) and its regionsomer 2.



Treatment of glycerol monobenzyl ether 3^3 with t-butyldimethylsilyl chloride (TBDMS-C]) under standard conditions⁴ gave a quantitative yield of the corresponding TBDMS ether 4 [b.p. 148-150° (3 mm)].⁵ Debenzylation of 4 was effected by hydrogenolysis under mild conditions (Scheme 1) giving the alcohol 5^6 in 98% yield. This reaction was sluggish in the absence of TBDMS-C1, which presumably is reduced to TBDMS-H and activates the catalyst.⁷ This debenzylation procedure is superior to that employing high pressure, which is known to cause silvl migration and desilvlation.⁸

The alcohol 5 served as the key intermediate for the construction of the C-12 side-chain (Scheme 1) ; a stepwise approach was selected because of its versatility in the synthesis of less unsaturated analogs of 1. To this end, Michael addition of the alcohol 5 to ethyl propiolate gave the unsaturated ester 6 in 74% yield. The E stereochemistry of the double bond in 6 was apparent from the 12 Hz coupling constants of its olefinic protons. Reduction of the ester 6 with DIBAL-H afforded the corresponding alcohol $\frac{7}{2}$ in quantitative yield. Oxidation of $\frac{7}{2}$ with manganese dioxide gave the unsaturated aldehyde 8 [δ 9.27 (1H, d, J = 9Hz, CHO), 7.23 (1H, d, J = 12 Hz, O-CH=C), 5.53 (1H, dd, J = 12 and 9 Hz, C=CH-CHO), 4.00-3.33 (5H, m, glycerol



SCHEME 2

CH), 0.80 (18 H, s, 2x SiC(CH₃)₃), 0.03 and 0.00 (6H each, s, 2x Si(CH₃)₂)] in 50% yield. The remainder of the product consisted chiefly of 5, probably arising from 8 due to a Michael attack of hydroxide ion present on the surface of MnO₂. Use of MnO₂ (pH = 8) prepared according to the method of Stork and Tomasz⁹ did not circumvent this decomposition which also occurred during the oxidation of 10 to 11.

Wittig-Horner reaction¹⁰ of the aldehyde <u>8</u> with the ylide prepared from diethyl carbethoxymethylphosphonate gave the diene ester <u>9</u> (44%) with E,E stereochemistry as deduced from its ¹H nmr spectrum [δ 7.20 (1H, dd, J = 15 and 12 Hz, Hc), 6.82 (1H, d, J = 12 Hz, Ha), 5.62 (1H, d, J = 15 Hz, Hd), 5.58 (1H, overlapping dd, J = 12 Hz, Hb), 4.10 (2H, q, J = 7.5 Hz, C0₂CH₂CH₃), 4.00-3.33 (5H, m, glycerol CH), 1.10 (3H, t, J = 7.5 Hz, C0₂CH₂CH₃), 0.83 (18H, 2x SiC(CH₃)₃) and 0.00 (12H, br s, 2x Si(CH₃)₂)]. DIBAL-H reduction of the ester <u>9</u> afforded the diene alcohol <u>10</u> (90%) which on oxidation with MnO₂ gave the diene aldehyde <u>11</u> (50% as estimated by ¹H nmr), contaminated with the decomposition product <u>5</u>. The aldehyde <u>11</u> without further purification was subjected to a Wittig reaction with the phosphorane derived from (E,E)-2,4-heptadienyltriphenylphosphonium bromide¹¹ to yield a mixture of E and Z pentaenes <u>12</u>. Deprotection of this mixture yielded a mixture of E and Z isomers of racemic fecapentaene-12 (<u>1</u>) which, like the natural product,¹ isomerized on standing to a mixture identical with an authentic sample as judged by TLC, co-HPLC, and UV.¹² Hydrogenation of <u>12</u> yielded a saturated derivative identical (TLC) with authentic material.

In connection with other studies we observed that treatment of the readily available monoacetin <u>13</u> with TBDMS-Cl under standard conditions yielded the 1,3-derivative <u>14</u>, after acetyl migration.¹³ Hydrolysis of <u>14</u> followed by Michael addition of the resulting alcohol <u>15</u> to propiolaldehyde afforded <u>16</u> with E stereochemistry as indicated by its ¹H nmr spectrum. Conversion of <u>16</u> to the pentaene <u>2</u> proceeded by the methods previously described (Scheme 2) and yielded a product isomeric with fecapentaene-12. The structure of <u>2</u> was confirmed by its ¹H nmr spectrum [$\delta(C_6D_6)$ 6.72 (1H, d, J = 12, -OCH=), 6.25-6.4 and 6.1-6.25 (8H, m, CH=CH), 5.75 (1H, m, OCH=CH), 3.4-3.9 (5H, m, glyceryl CH) 2.08 (2H, dq, J ~ 8Hz, =CHCH_2CH_3), 1.00 (3H, t, -CH₂CH₃)], and by hydrogenation of its protected derivative <u>18</u> to the saturated analog and comparison with an authentic sample (TLC). Compound <u>2</u> was found to be as mutagenic in the Ames test as fecapentaene-12.¹²

An interesting observation was made whilst attempting to find optimum conditions for the Michael reaction of alcohol 5 with methyl propiolate. Use of triethylamine in THF at 0°C for 1 hr yielded a mixture of the expected adduct 6 and its isomer 17. Apparently, silyl migration of 5 to give the isomer 15 occurred at a rate faster than Michael addition; presumably relief of steric crowding provides the driving force for this unusual reaction.

The availability of fecapentacene-12 by this route will make it possible for us to explore further its role in the etiology of human colon cancer.

<u>Acknowledgement</u>. It is a pleasure to acknowledge the support of this investigation by the National Cancer Institute through Grant CA-23857.

REFERENCES AND NOTES

- Hirai, N.; Kingston, D.G.I.; Van Tassell, R. L.; Wilkins, T. D. J. Amer. Chem. Soc. 1982, 104, 6149.
- Bruce, W. R.; Baptista, J.; Che, T.; Furrer, R.; Gingerich, J. D.; Gupta, I.; Krepinsky, J. J.; Grey, A. A.; Yates, P. <u>Naturwiss 1982, 69</u>, 557. Gupta, I.; Baptista, J.; Bruce, W. R.; Che, C. T.; Furrer, R.; Gingerich, J. S.; Grey, A. A.; Marai, L.; Yates, P.; Krepinsky, J. J. Biochemistry 1983, 22, 241.
- 3. Sowden, J. C.; Fischer, H. O. L. J. Amer. Chem. Soc. 1941, 63, 3244.
- 4. Corey, E. J.; Venkateswarlu, A. J. Amer. Chem. Soc. 1972, 94, 6190.
- 5. Structural assignments of all new compounds are based on i.r., 1 H n.m.r. in CDCl3, and, in most cases, mass spectroscopic evidence.
- 6. Compounds 5-12 (scheme 1) and 14-17 (scheme 2) are all obtained as nondistillable oils.
- 7. Emerson, G. F.; Pettit, R. J. Amer. Chem. Soc. 1962, 84, 4591.
- 8. Dodd, G. H.; Golding, B. T.; Ioannou, P. V. J. Chem. Soc., Chem. Commun 1975, 249.
- 9. Stork, G.; Tomasz, M. J. Amer. Chem. Soc. 1964, 86, 471.
- 10. Pettit, G. R.; Knight, J. C.; Herald, C. L. J. Org. Chem. 1970, 35, 1393.
- Obtained from (E,E)-2,4-heptadienol prepared by the reduction of commercially available (E,E)-2,4-heptadienal.
- 12. We thank Mr. Roger Van Tassell for this determination.
- 13. Hibbert, H.; Carter, N. M. J. Amer. Chem. Soc. 1929, 51, 1601.

(Received in USA 15 September 1983)